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	Terms	Documents			
	L1 same (organophosphate or malathion)	29			
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<u>L2</u>	L1 same (organophosphate or malathion)	29	<u>L2</u>
<u>L1</u>	carboxylesterase	347	<u>L1</u>

END OF SEARCH HISTORY

## WEST

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## Search Results - Record(s) 1 through 29 of 29 returned.

L2: Entry 1 of 29

File: PGPB

May 15, 2003

PGPUB-DOCUMENT-NUMBER: 20030091975

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20030091975 A1

TITLE: Multiple determinants for metabolic phenotypes

PUBLICATION-DATE: May 15, 2003

INVENTOR-INFORMATION:

NAME

CITY

STATE

COUNTRY

RULE-47

Leyland-Jones, Brian

Miami

FL

US

US-CL-CURRENT: 435/4; 424/9.2

## Full Title Citation Front Review Classification Date Reference Sequences Attachments Claims KMC Draw Desc Image

\_\_\_\_\_ 2. Document ID: US 20030077222 A1

L2: Entry 2 of 29

File: PGPB

Apr 24, 2003

PGPUB-DOCUMENT-NUMBER: 20030077222

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20030077222 A1

TITLE: Individualization of therapy with analgesics

PUBLICATION-DATE: April 24, 2003

INVENTOR-INFORMATION:

NAME

CITY

STATE

COUNTRY

RULE-47

Leyland-Jones, Brian

Miami

FL

US

US-CL-CURRENT: 424/9.1; 435/7.92

Full Title Citation Front Remem Classification Date Reference Sequences Attachments Claims Mill Draw Desc Image

☐ 3. Document ID: US 20030073133 A1

L2: Entry 3 of 29

File: PGPB

Apr 17, 2003

PGPUB-DOCUMENT-NUMBER: 20030073133

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20030073133 A1

TITLE: Individualization of therapy with erectile dysfunction agents

PUBLICATION-DATE: April 17, 2003

INVENTOR - INFORMATION:

NAME

CITY STATE

COUNTRY RULE-47

Leyland-Jones, Brian

Miami

FL

US-CL-CURRENT: 435/7.1

Full Title Offation Front Review Classification Date Reference Sequences Attachments Claims Milit Draw Desc Image

☐ 4. Document ID: US 20030072710 A1

L2: Entry 4 of 29

File: PGPB

Apr 17, 2003

PGPUB-DOCUMENT-NUMBER: 20030072710

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20030072710 A1

TITLE: Individualization of therapy with antidepressants

PUBLICATION-DATE: April 17, 2003

INVENTOR-INFORMATION:

NAME

CITY

STATE

COUNTRY

RULE-47

Leyland-Jones, Brian

Miami

Full Title Citation Front Review Classification Date Reference Sequences Attachments Claims (Will Draw Desc Image)

FL

US-CL-CURRENT: 424/9.1; 424/9.2

L2: Entry 5 of 29

File: PGPB

Apr 10, 2003

PGPUB-DOCUMENT-NUMBER: 20030068273

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20030068273 A1

TITLE: Individualization of therapy with immunosuppressants

PUBLICATION-DATE: April 10, 2003

INVENTOR-INFORMATION:

NAME

CITY

STATE

COUNTRY

RULE-47

Leyland-Jones, Brian

Miami

 ${ t FL}$ 

US

US-CL-CURRENT: 424/9.2

Full Title Citation Front Review Classification Date Reference Sequences Attachments

MMC Draw Desc Image

☐ 6. Document ID: US 20030053950 A1

L2: Entry 6 of 29

File: PGPB

Mar 20, 2003

PGPUB-DOCUMENT-NUMBER: 20030053950

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20030053950 A1

TITLE: Individualization of therapy with hyperlipidemia agents

PUBLICATION-DATE: March 20, 2003

INVENTOR - INFORMATION:

NAME

CITY

STATE

COUNTRY

RULE-47

Leyland-Jones, Brian

Miami

FL

US

US-CL-CURRENT: 424/9.1; 435/7.1

Full Title Otation Front Review Classification Date Reference Sequences Attachments

K000C - Draw Desc - Image

7. Document ID: US 20030049204 A1

L2: Entry 7 of 29

File: PGPB

Mar 13, 2003

PGPUB-DOCUMENT-NUMBER: 20030049204

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20030049204 A1

TITLE: Individualization of therapy with gastroesophageal reflux disease agents

PUBLICATION-DATE: March 13, 2003

INVENTOR - INFORMATION:

NAME

CITY

STATE COUNTRY

RULE-47

Leyland-Jones, Brian

Miami

 $\mathtt{FL}$ 

US

US-CL-CURRENT: 424/9.1; 435/7.92

Full Title Offation Front Review Classification Date Reference Sequences Attachments

\_\_\_\_\_\_ 8. Document ID: US 20020184655 A1

L2: Entry 8 of 29

File: PGPB

Dec 5, 2002

PGPUB-DOCUMENT-NUMBER: 20020184655

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20020184655 A1

TITLE: METHODS FOR THE DEGRADATION AND DETOXIFICATION OF ORGANIC MATERIAL USING URINE PRODUCED BY TRANSGENIC ANIMALS AND RELATED TRANSGENIC ANIMALS AND PROTEINS

PUBLICATION-DATE: December 5, 2002

INVENTOR - INFORMATION:

NAME LUBON, HENRYK PALEYANDA, REKHA DROHAN, WILLIAM VELANDER, WILLIAM

CITY ROCKVILLE GAITHERSBURG

SPRINGFIELD

BLACKSBURG

COUNTRY STATE MD US MD US VA US VA US

RULE-47

US-CL-CURRENT: 800/4; 800/14, 800/18, 800/24, 800/8

Full Title Citation Front Review Classification Date Reference Sequences Attachments

MildC | Draw Desc | Image

1 9. Document ID: US 20020182636 A1

L2: Entry 9 of 29

File: PGPB

Dec 5, 2002

PGPUB-DOCUMENT-NUMBER: 20020182636

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20020182636 A1

TITLE: 53010, a novel human carboxylesterase family member and uses thereof

PUBLICATION-DATE: December 5, 2002

INVENTOR - INFORMATION:

NAME

CITY

STATE COUNTRY

RULE-47

Curtis, Rory A. J.

Southborough

US MΑ

Silos-Santiago, Inmaculada Jamaica Plain

US

US-CL-CURRENT: 435/7.1; 435/196, 435/320.1, 435/325, 435/69.1, 530/388.26, 536/23.2

Full Title Citation Front Review Classification Date Reference Sequences Attachments

PMC | Draw Desc | Image

☐ 10. Document ID: US 20020168713 A1

L2: Entry 10 of 29

File: PGPB

Nov 14, 2002

PGPUB-DOCUMENT-NUMBER: 20020168713

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20020168713 A1

TITLE: 46980, a novel human neuroligin family member and uses thereof

PUBLICATION-DATE: November 14, 2002

INVENTOR-INFORMATION:

NAME

CITY

STATE

COUNTRY

US

RULE-47

Curtis, Rory A.J.

Southborough

MA

Full Title Citation Front Review Classification Date Reference Sequences Attachments

US-CL-CURRENT: 435/69.1; 435/320.1, 435/325, 536/23.2

kinic Draw Desc Image

11. Document ID: US 20020151068 A1

L2: Entry 11 of 29

File: PGPB

Oct 17, 2002

PGPUB-DOCUMENT-NUMBER: 20020151068

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20020151068 A1

TITLE: Compositions and methods for the diagnosis and treatment of organophosphate toxicity

PUBLICATION-DATE: October 17, 2002

INVENTOR - INFORMATION:

NAME CITY STATE COUNTRY RULE-47

Haley, Robert Dallas TX US Varley, Alan Plano TX US Munford, Robert Dallas TX US

US-CL-CURRENT: 435/456; 435/320.1, 514/44

Full Title Citation Front Review Classification Date Reference Sequences Attachments MIMC Draw Desc Image

☐ 12. Document ID: US 20020150910 A1

L2: Entry 12 of 29

File: PGPB

Oct 17, 2002

PGPUB-DOCUMENT-NUMBER: 20020150910

PGPUB-FILING-TYPE: new

DOCUMENT-IDENTIFIER: US 20020150910 A1

TITLE: 33410, a novel human carboxylesterase family member and uses thereof

PUBLICATION-DATE: October 17, 2002

INVENTOR - INFORMATION:

NAME

CITY

STATE

COUNTRY

RULE-47

Curtis, Rory A.J.

Southborough

MA

US

US-CL-CURRENT:  $\frac{435}{6}$ ;  $\frac{435}{196}$ ,  $\frac{435}{320.1}$ ,  $\frac{435}{325}$ ,  $\frac{435}{69.1}$ ,  $\frac{435}{7.1}$ ,  $\frac{530}{388.26}$ ,  $\frac{536}{23.2}$ 

Full Title Citation Front Review Classification Date Reference Sequences Attachments

KNMC Draw Desc Image

☐ 13. Document ID: US 6436437 B1

L2: Entry 13 of 29

File: USPT

Aug 20, 2002

US-PAT-NO: 6436437

DOCUMENT-IDENTIFIER: US 6436437 B1

TITLE: Covalent polar lipid conjugates with neurologically active compounds for targeting

Full Title Citation Front Review Classification Date Reference Sequences Attachments

R000C - Oraxo Desc - Image |

☐ 14. Document ID: US 6387876 B1

L2: Entry 14 of 29

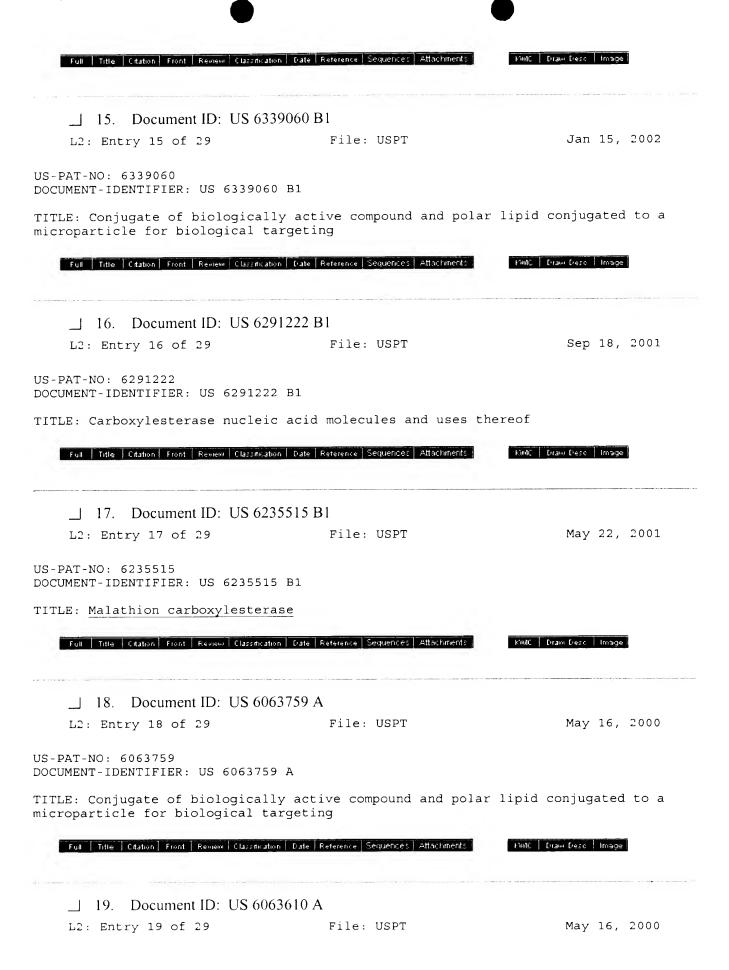
File: USPT

May 14, 2002

US-PAT-NO: 6387876

DOCUMENT-IDENTIFIER: US 6387876 B1

TITLE: Covalent polar lipid-conjugates with biologically active compounds for use in salves



US-PAT-NO: 6063610

DOCUMENT-IDENTIFIER: US 6063610 A

TITLE: Carboxylesterase nucleic acid molecules, proteins and uses thereof

Full Title Citation Front Review Classification Date Reference Sequences Attachments

PMC Draw Desc Image

1 20. Document ID: US 6024977 A

L2: Entry 20 of 29

File: USPT

Feb 15, 2000

US-PAT-NO: 6024977

DOCUMENT-IDENTIFIER: US 6024977 A

TITLE: Covalent polar lipid conjugates with neurologically active compounds for

targeting

Full Title Citation Front Review Classification Date Reference Sequences Attachments

KMC Draw Desc Image

L2: Entry 21 of 29

File: USPT

Dec 14, 1999

US-PAT-NO: 6001625

DOCUMENT-IDENTIFIER: US 6001625 A

TITLE: Site-directed mutagenesis of esterases

Full Title Citation Front Review Classification Date Reference Sequences Attachments

KMMC | Draw Desc | Image

□ 22. Document ID: US 5965519 A

L2: Entry 22 of 29

File: USPT

Oct 12, 1999

US-PAT-NO: 5965519

DOCUMENT-IDENTIFIER: US 5965519 A

TITLE: Covalent polar lipid conjugates with biologically-active compounds for use in

salves

Full Title Citation Front Review Classification Date Reference Sequences Attachments

PMC Draw Desc Image

L2: Entry 23 of 29

File: USPT

Dec 1, 1998

US-PAT-NO: 5843758

DOCUMENT-IDENTIFIER: US 5843758 A

TITLE: Enzyme based bioremediation

Full Title Citation Front Review Classification Date Reference Sequences Attachments

Finit Draw Desc Image

☐ 24. Document ID: US 5827819 A

L2: Entry 24 of 29

File: USPT

Oct 27, 1998

US-PAT-NO: 5827819

DOCUMENT-IDENTIFIER: US 5827819 A

TITLE: Covalent polar lipid conjugates with neurologically active compounds for

targeting

Full Title Citation Front Review Classification Date Reference Sequences Attachments

KWWC Draw Desc Image

☐ 25. Document ID: US 5716831 A

L2: Entry 25 of 29

File: USPT

Feb 10, 1998

US-PAT-NO: 5716831

DOCUMENT-IDENTIFIER: US 5716831 A

TITLE: Method and test kit for detecting insecticide resistance

Full Title Citation Front Review Classification Date Reference Sequences Attachments

KMC | Draw Desc | Image |

L2: Entry 26 of 29

File: USPT

Dec 20, 1977

US-PAT-NO: 4064237

DOCUMENT-IDENTIFIER: US 4064237 A

\*\* See image for Certificate of Correction \*\*

TITLE: Synergistic pesticidal mixtures of phosalone and malathion and process for controlling arthropods therewith

5

☐ 27. Document ID: WO 9719176 A1

L2: Entry 27 of 29

File: EPAB

May 29, 1997

PUB-NO: WO009719176A1

DOCUMENT-IDENTIFIER: WO 9719176 A1 TITLE: MALATHION CARBOXYLESTERASE

Full Title Citation Front Review Classification Date Reference Sequences Attachments

Full Title Citation Front Review Classification Date Reference Sequences Attachments

KMMC | Drawi Desc | Image

KMMC | Draw Desc | Image |

L2: Entry 28 of 29

File: DWPI

Dec 14, 1999

DERWENT-ACC-NO: 2000-096137

DERWENT-WEEK: 200008

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TITLE: Enhancing the organophosphate detoxifying capabilities of esterases for the treatment of organophosphate poisoning

and a second		15 B1 AU 9675572 A ZA 9609824 A A MX 9804053 A1 JP 2000504203 W
L2: Entry 29 of 29	File: DWPI	May 29, 199
NT-ACC-NO: 1997-298113 NT-WEEK: 200130 IGHT 2003 DERWENT INFO		
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tamination of soil, wa	ater, food etc	Attachments FMC Draw Desc Image

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=> d 15 ibib ab 1-13 ANSWER 1 OF 13 SCISEARCH COPYRIGHT 2003 THOMSON ISI 2002:831160 SCISEARCH ACCESSION NUMBER: THE GENUINE ARTICLE: 599XW Purification and characterization of a TITLE: carboxylesterase involved in malathion-specific resistance from Tribolium castaneum (Coleoptera : Tenebrionidae) Haubruge E (Reprint); Amichot M; Cuany A; Berge J B; AUTHOR: Arnaud L Gembloux Agr Univ, Dept Pure & Appl Zool, B-5030 Gembloux, CORPORATE SOURCE: Belgium (Reprint); INRA, Lab Biol Invertebres, Ctr Antibes, F-06606 Antibes, France Belgium; France COUNTRY OF AUTHOR: INSECT BIOCHEMISTRY AND MOLECULAR BIOLOGY, (SEP 2002) Vol. SOURCE: 32, No. 9, pp. 1181-1190. Publisher: PERGAMON-ELSEVIER SCIENCE LTD, THE BOULEVARD, LANGFORD LANE, KIDLINGTON, OXFORD OX5 1GB, ENGLAND. ISSN: 0965-1748. Article; Journal DOCUMENT TYPE: LANGUAGE: English REFERENCE COUNT: 40 \*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\* Specific resistance to malathion in a strain of Tribolium castaneum is AB due to a 44-fold increase in malathion carboxylesterase (MCE) activity relative to a susceptible strain, whereas non-specific esterase levels are slightly lower. Unlike the overproduced esterase of some mosquito and aphid species, MCE in Tribolium castaneum accounts for only a small fraction (0.033-0.045%) of the total extractable protein respectively in resistant and susceptible strains. The enzyme was purified to apparent homogeneity from these two strains and has a similar molecular weight of 62,000. However, preparative isoelectricfocusing indicated that resistant insects possess one MCE with pI of 7.3, while susceptible insects possess a MCE with a pI of 6.6. Purified MCE from both populations had different K-m and V-m values for hydrolysis of malathion as well as for alpha-naphthyl acetate. The kinetic analysis suggests that MCE of resistant insects hydrolyses malathion faster than the purified carboxylesterase from susceptible beetles and that this enzyme has greater affinity for malathion than for naphthyl esters. Malathion-specific resistance is due to the presence of a qualitatively different esterase in the resistant

ANSWER 2 OF 13 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC.DUPLICATE L5 1

2000:187424 BIOSIS ACCESSION NUMBER: DOCUMENT NUMBER: PREV200000187424

MCE activities and malathion resistances in field TITLE:

strain. (C) 2002 Elsevier Science Ltd. All rights reserved.

populations of the Australian sheep blowfly (Lucilia

cuprina.

Smyth, Kerrie-Ann (1); Boyce, Thomas M.; Russell, Robyn J.; AUTHOR(S):

Oakeshott, John G.

CORPORATE SOURCE: (1) Institute of Cellular and Molecular Biology, University

of Texas at Austin, Molecular Biology Building, Austin, TX,

78712-1095 USA

Heredity, (Jan., 2000) Vol. 84, No. 1, pp. 63-72. SOURCE:

ISSN: 0018-067X.

DOCUMENT TYPE: Article LANGUAGE: English SUMMARY LANGUAGE: English

Malathion resistance has been shown to be result of a single point mutation in the LcalphaE7 gene in four independently isolated chromosomes of Lucilia cuprina. The resultant amino acid substitution specifies high malathion carboxylesterase

(MCE) activity. We have assayed MCE activities and resistance to malathion in three sets of field-derived samples, two sets of isogenic lines and five mass populations, and show that resistance to malathion in these samples is associated with high MCE activity in both sets of isogenic lines and four of the five mass populations. Additional mechanisms contributing to MCE activity or malathion resistance may be present in one of the mass populations. A second point mutation in LcalphaE7 is responsible for conferring diazinon resistance by encoding an increased organophosphate (OP) hydrolase activity. We also assayed diazinon resistances from the same three samples and show that diazinon and malathion resistances were in complete disequilibrium, with two exceptions. One exception involves the mass population with additional resistance mechanism(s) and the other involves three isogenic lines that are resistant to both insecticides. The molecular data for these lines suggest that they carry a duplication of the LcalphaE7 gene.

L5 ANSWER 3 OF 13 SCISEARCH COPYRIGHT 2003 THOMSON ISI

ACCESSION NUMBER: 1999:56480 SCISEARCH

THE GENUINE ARTICLE: 154CR

TITLE: Characterization of esterases in

malathion-resistant and susceptible strains of the pteromalid parasitoid Anisopteromalus calandrae

AUTHOR: Baker J E (Reprint); Fabrick J A; Zhu K Y

CORPORATE SOURCE: USDA ARS, GRAIN MKT PROD & RES CTR, 1515 COLL AVE,

MANHATTAN, KS 66502 (Reprint); KANSAS STATE UNIV, DEPT

ENTOMOL, MANHATTAN, KS 66506

COUNTRY OF AUTHOR: USA

SOURCE:

INSECT BIOCHEMISTRY AND MOLECULAR BIOLOGY, (DEC 1998) Vol.

28, No. 12, pp. 1039-1050.

Publisher: PERGAMON-ELSEVIER SCIENCE LTD, THE BOULEVARD,

LANGFORD LANE, KIDLINGTON, OXFORD OX5 1GB, ENGLAND.

ISSN: 0965-1748.

DOCUMENT TYPE:

Article; Journal

FILE SEGMENT:

LIFE; AGRI

LANGUAGE:

English

REFERENCE COUNT: 71

\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

General esterase, malathion-specific carboxylesterase, AB phosphotriesterase, glutathione S-transferase, cytochrome P-450-dependent monooxygenase activity, and target site sensitivity were compared in malathion-resistant (R) and malathion-susceptible (S) strains of the parasitoid Anisopteromalus calandrae (Howard) (Hymenoptera: Pteromalidae). Activity against alpha-naphthyl acetate was not significantly different in male and female wasps for either strain. General esterase activity ranged from 1.2-fold to 2.5-fold higher in the R strain compared with the S strain, but these differences between strains were not consistent. Based on V-max/K-m ratios estimated for a number of analogs of four substrates (alpha-naphthyl acetate, beta-naphthyl acetate, 4-methylumbelliferyl acetate, and p-nitrophenyl acetate) there was no evidence that general esterase activity was elevated or reduced in the R strain. Malathion-specific carboxylesterase (MCE) activity, determined by using 2,3-C-14-malathion as substrate, was 10- to 30-fold higher in the R strain compared with that in the S strain. The MCE. has a pH optima at about pH 7, is cytosolic, and is labile upon storage at - 80 degrees C. MCE activity could be recovered from native 10% PAGE gels and IEF-PAGE gels (pI = 5.2), but the peak of MCE activity also contained the major peak of activity against cy-naphthyl acetate. There was no evidence for major involvement of phosphotriesterase, glutathione S-transferase, monooxygenase, or altered acetylcholinesterase in the resistance. These data suggest that an increased activity of a MCE in the R strain is the probable major mechanism conferring resistance to malathion in A. calandrae. This study provides the first characterization of a biochemical resistance mechanism in a parasitoid with a high level of resistance to an organophosphate insecticide. (C) 1998 Elsevier Science

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L5 ANSWER 4 OF 13 SCISEARCH COPYRIGHT 2003 THOMSON ISI

ACCESSION NUMBER: 1998:362837 SCISEARCH

THE GENUINE ARTICLE: ZL902

TITLE .

Cross-resistance patterns among Lucilia cuprina

(Diptera: Calliphoridae) resistant to organophosphorus

insecticides

AUTHOR: Campbell P M (Reprint); Yen J L; Masoumi A; Russell R J;

Batterham P; McKenzie J A; Oakeshott J G

CORPORATE SOURCE: CSIRO, DIV ENTOMOL, POB 1700, CANBERRA, ACT 2601,

AUSTRALIA (Reprint)

COUNTRY OF AUTHOR:

SOURCE:

AUSTRALIA
JOURNAL OF ECONOMIC ENTOMOLOGY, (APR 1998) Vol. 91, No. 2,

pp. 367-375.

Publisher: ENTOMOL SOC AMER, 9301 ANNAPOLIS RD, LANHAM, MD

20706.

ISSN: 0022-0493. Article; Journal

DOCUMENT TYPE: FILE SEGMENT:

AGRI

LANGUAGE:

English

REFERENCE COUNT: 43

\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

Strains of Lucilia cuprina (Wiedemann) have been AR characterized as having low, intermediate, or high levels of esterase-mediated hydrolysis Of the organophosphorus insecticide, chlorfenvinphos. These levels correlate respectively with susceptibility to organophosphorus insecticides, malathion resistance, or diazinon resistance. Diazinon and chlorfenvinphos are diethyl organophosphorus insecticides having 2 ethoxy groups attached to their central phosphorus atom, whereas malathion is a dimethyl organophosphorus insecticide having 2 methoxy groups attached to its phosphorus atom, and, unusually, malathion also has 2 carboxylester bonds in addition to the phosphoester bonds that define organophosphorus compounds. We tested larvae for resistance to diazinon and also assessed representative malathion-resistant and diazinon-resistant L. cuprina strains at the adult stage for resistance to 12 organophosphorus insecticides, including analog pairs differing only in respect to their dimethyl-diethyl status. Two malathion-resistant strains have low-level cross-resistance to diazinon (3 to 4-fold), 4 diazinon-resistant strains have high-level diazinon resistance (11 to 16-fold), and 2 strains with a combined (malathion plus diazinon) resistance type also have high-level diazinon resistance (17 to 18-fold) relative to 3 organophosphorus insecticide-susceptible strains. One of the diazinon-resistant strains showed approximate to 2 times greater resistance factors toward diethyl organophosphorus insecticides than their dimethyl analogs while (leaving aside malathion to consider only the majority which have no carboxylester groups) a malathion-resistant strain showed 2-5 times greater resistance factors toward the dimethyl organophosphorus insectides than their diethyl analogs. The diazinon-resistant strain showed no resistance to 2 di-isopropyl organophosphorus compounds or to 2 organophosphorus insecticides which are asymmetric about the phosphorus atom (optically active). The malathion-resistant strain showed only slight resistance (<3-fold) to either the di-isopropyl or optically active organophosphorus insecticides, including the di-isobropyl analog of malathion. These cross-resistance patterns parallel those of certain organophosphorus insecticide-resistant strains of Musca domestica L., in which diazinon and malathion resistances also are proposed to be esterase mediated, reinforcing other biochemical data suggesting a general mechanism among the higher Diptera.

L5 ANSWER 5 OF 13 SCISEARCH COPYRIGHT 2003 THOMSON ISI

ACCESSION NUMBER: 1998:422333 SCISEARCH

THE GENUINE ARTICLE: ZQ240

TITLE: Propetamphos resistance in the Australian sheep blowfly,

Lucilia cuprina (Wiedemann) (Diptera:

Calliphoridae)

AUTHOR: Smyth K A (Reprint); Russell R J; Oakeshott J G

CORPORATE SOURCE: SYRACUSE UNIV, DEPT BIOL, BIOL RES LABS, SYRACUSE, NY

13244 (Reprint); CSIRO, DIV ENTOMOL, CANBERRA, ACT 2601,

AUSTRALIA

COUNTRY OF AUTHOR: USA;

USA; AUSTRALIA

SOURCE:

AUSTRALIAN JOURNAL OF ENTOMOLOGY, (3 APR 1998) Vol. 37,

Part 1, pp. 57-59.

Publisher: BLACKWELL SCIENCE, 54 UNIVERSITY ST, P O BOX

378, CARLTON VICTORIA 3053, AUSTRALIA.

ISSN: 1326-6756.

DOCUMENT TYPE:

Article; Journal

FILE SEGMENT: LANGUAGE: AGRI English

REFERENCE COUNT:

English 20

\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

AB Lucilia cuprina lines, previously characterised by their resistance to diazinon and malathion, were tested for their resistance to another organophosphate, propetamphos. All 13 lines tested showed no difference in propetamphos tolerance, regardless of their resistance to diazinon, malathion or both. It is concluded that resistance to one structural type of organophosphate does not necessarily confer

L5 ANSWER 6 OF 13 CAPLUS COPYRIGHT 2003 ACS

resistance to another.

ACCESSION NUMBER:

1997:450131 CAPLUS

DOCUMENT NUMBER:

127:77923

TITLE:

Malathion carboxylesterases of

resistant Lucilia cuprina for bioremediation

of insecticide contamination

INVENTOR(S):

Russell, Robyn Joyce; Newcomb, Richard David;

Campbell, Peter Malcolm; Robin, Geoffrey Charles De Quetteville; Claudianos, Charles; Smyth, Kerrie-ann; Boyce, Thomas Mark; Oakeshott, John Graham; Brownlie,

Jeremy Colin; et al.

PATENT ASSIGNEE(S):

Commonwealth Scientific and Industrial Research Organisation, Australia; Russell, Robyn Joyce;

Organisation, Australia; Russell, Robyn Joyce; Newcomb, Richard David; Campbell, Peter Malcolm

SOURCE:

PCT Int. Appl., 56 pp.

DOCUMENT TYPE:

CODEN: PIXXD2 Patent

LANGUAGE:

English

FAMILY ACC. NUM. COUNT:

1

PATENT INFORMATION:

PATENT NO.	KIND DATE	APPLICATION NO. DATE
wo 9719176	1 19970529	WO 1996-AU746 19961122
W: AL, AM,	AT, AU, AZ, BA, BB,	BG, BR, BY, CA, CH, CN, CU, CZ, DE,
DK, EE,	ES, FI, GB, GE, HU,	IL, IS, JP, KE, KG, KP, KR, KZ, LC,
LK, LR,	LS, LT, LU, LV, MD,	MG, MK, MN, MW, MX, NO, NZ, PL, PT,
RO, RU,	SD, SE, SG, SI, SK,	TJ, TM, TR, TT, UA, UG, US, UZ, VN,
AM, AZ,	BY, KG, KZ, MD, RU,	TJ, TM
RW: KE, LS,	MW, SD, SZ, UG, AT,	BE, CH, DE, DK, ES, FI, FR, GB, GR,
IE, IT,	LU, MC, NL, PT, SE,	BF, BJ, CF, CG, CI, CM, GA, GN, ML,
MR, NE,	SN, TD, TG	
CA 2236793	AA 19970529	CA 1996-2236793 19961122
AU 9675572	A1 19970611	AU 1996-75572 19961122
AU 700336	B2 19981224	
ZA 9609824	A 19970708	ZA 1996-9824 19961122
EP 862636	A1 19980909	EP 1996-937941 19961122
R: AT, BE,	CH, DE, DK, ES, FR,	GB, GR, IT, LI, LU, NL, SE, MC, PT,

IE, FI BR 1996-11627 19961122 19990406 BR 9611627 Α 20000411 JP 1997-519237 19961122 JP 2000504203 T2 20010522 US 1998-68960 19980520 US 6235515 В1 A 19951123 AU 1995-6751 PRIORITY APPLN. INFO.: WO 1996-AU746 W 19961122 Genes and cDNAs encoding malathion carboxylesterases AB of the sheep blowfly (Lucilia cuprina) that are capable of hydrolyzing at least one organophosphate selected from the group consisting of carboxylester organophosphates and dimethyloxon organophosphates are described. Genes encoding several isoenzymes are identified and the enzymes characterized and the preferred enzymes are variants of the isoenzyme encoded by the Lc.alpha.E7 gene. The preferred analogs have an amino acid substitution of Trp-251 selected from the group consisting of Leu, Ser, Ala, Ile, Val, Thr, Cys, Met and Gly. The preferred substituents are Leu and Ser. These substitutions were identified by sequencing of a no. of cloned genes from Lucilia and the orthologous enzyme from Musca domestica. ANSWER 7 OF 13 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC.DUPLICATE ACCESSION NUMBER: 1997:391285 BIOSIS DOCUMENT NUMBER: PREV199799690488 Isolation and characterization of an TITLE: unamplified esterase B3 gene from malathion-resistant Culex tarsalis. Tittiger, Claus; Walker, Virginia K. (1) AUTHOR(S): (1) Dep. Biol., Queen's Univ., Kingston, ON K7L 3N6 Canada CORPORATE SOURCE: Biochemical Genetics, (1997) Vol. 35, No. 3-4, pp. 119-138. SOURCE: ISSN: 0006-2928. DOCUMENT TYPE: Article LANGUAGE: English A malathion-resistant strain of Culex tarsalis has a malathion carboxylesterase which rapidly hydrolyzes the insecticide. This is in contrast to organophosphate-resistant strains of C. quinquefasciatus and C. pipiens, which have elevated levels of general B esterases due to amplification of the corresponding genes, producing increased amounts of enzyme which appear to protect the insects by sequestering the insecticide. The contribution to resistance of the homologous esterase B3 (Est-beta-3) gene (est-beta-3) in C. tarsalis was investigated by cloning and characterizing sequences from resistant and susceptible strains. est-beta-3 is similar to est-beta-1, both structurally and in sequence. The first intron of est-beta-3, however has a region of extensive repeats which may be responsible for the inefficient processing of the transcript. Southern blots indicate that the gene is single copy in both strains, and northern blots show that it is not greatly overexpressed in the resistant insects. est-beta-3 cDNAs from resistant and susceptible strains have 98% amino acid identity. It appears that, in contrast to other studies, est-beta-3 does not play a significant role in insecticide resistance in our strains of C. tarsalis, and the molecular responses of pest insects to organophosphates may be more diverse than has been suggested. ANSWER 8 OF 13 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC.DUPLICATE 1995:413994 BIOSIS ACCESSION NUMBER: DOCUMENT NUMBER: PREV199598428294 Characterization of a novel esterase conferring TITLE: insecticide resistance in the mosquito Culex tarsalis. Whyard, Steven; Downe, Aylward E. R.; Walker, Virginia K. AUTHOR(S): (1) (1) Dep. Biol., Queen's Univ., Kingston, ON K7L 3N6 Canada CORPORATE SOURCE: Archives of Insect Biochemistry and Physiology, (1995) Vol. SOURCE:

29, No. 4, pp. 329-342.

ISSN: 0739-4462.

DOCUMENT TYPE: LANGUAGE:

Article English

Resistance to the organophosphate insecticide, malathion, in a strain of Culex tarsalis mosquitoes is due to increased activity of a malathion carboxylesterase (MCE). To determine whether resistance was due to a qualitative or quantitative change in the MCE, the enzyme was purified from both malathion-resistant and -susceptible mosquitoes. Enzyme kinetic measurements revealed that the two strains have one MCE in common, but resistant mosquitoes also have a unique MCE which hydrolyses malathion 18 times faster. Interestingly, this MCE does not hydrolyse alpha-naphthyl acetate, a substrate commonly used to detect increased levels of esterases in other organophosphate-resistant insects. Unlike the over-produced esterase of some related mosquito species, each MCE in C. tarsalis accounts for only a small fraction (0.015%) of the total extractable protein in either strain.

Therefore, resistance in these insects is due to the presence of a qualitatively different enzyme, and not to a quantitative increase of a non-specific esterase. This study therefore demonstrates that the underlying biochemical mechanisms of insecticide resistance in one insect cannot necessarily be predicted from those of another, even closely related species.

ANSWER 9 OF 13 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC.DUPLICATE

ACCESSION NUMBER:

1994:500223 BIOSIS PREV199497513223

DOCUMENT NUMBER: TITLE:

Isolation of an esterase conferring insecticide

resistance in the mosquito Culex tarsalis.

AUTHOR(S):

Whyard, S.; Downe, A. E. R.; Walker, V. K. (1)

CORPORATE SOURCE:

(1) Dep. Biol. Insect Biotech Canada, Queen's Univ.,

Kingston, ON K7L 3N6 Canada

SOURCE:

Insect Biochemistry and Molecular Biology, (1994) Vol. 24,

No. 8, pp. 819-827.

ISSN: 0965-1748.

DOCUMENT TYPE:

Article English LANGUAGE:

Malathion resistance in a strain of Culex tarsalis mosquitoes is due primarily to the activity of a malathion carboxylesterase (MCE). The resistant strain was 150 times more resistant to malathion than the susceptible strain and was weakly resistant to malaoxon and carbaryl, but not to any other insecticide tested. The phenotype could be reversed with the carboxylesterase inhibitor triphenylphosphate, but no synergism was observed with either the phosphatase or polysubstrate monooxygenase inhibitors, NaF and piperonyl butoxide. MCE is expressed throughout development and is most concentrated in the gut tissues of the larvae. Subcellular fractionation indicated that MCE was localized primarily in the mitochondria of resistant insects and the cytoplasm of susceptible insects. The enzyme was purified to homogeneity from both strains. and has a molecular weight of 59,000. However, chromatofocusing indicated that resistant insects have two MCEs with pIs of 6.8 and 6.2, while susceptible insects possessed only one MCE with a pI of 6.8. The MCE unique to the resistant strain hydrolysed malathion 18 times faster than the MCE common to both strains, suggesting that malathion resistance in C. tarsalis is due to the presence of a qualitatively different esterase in the resistant strain.

ANSWER 10 OF 13 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC.DUPLICATE

ACCESSION NUMBER: 1995:109131 BIOSIS DOCUMENT NUMBER: PREV199598123431

TITLE: Characterization of malathion carboxylesterase in the sheep blowfly Lucilia cuprina.

Whyard, Steven; Walker, Virginia K. (1) AUTHOR (S):

(1) Dep. Biol. Insect Biotech Canada, Queen's Univ., CORPORATE SOURCE:

Kington, ON K7L 3N6 Canada

Pesticide Biochemistry and Physiology, (1994) Vol. 50, No. SOURCE:

3, pp. 198-206. ISSN: 0048-3575.

Article DOCUMENT TYPE: English LANGUAGE:

Resistance to malathion in a strain of the Australian sheep blowfly is due to a 10-fold increase in malathion carboxylesterase

(MCE) activity relative to a more susceptible strain. MCE was purified to apparent homogeneity from these two strains and was shown to be a monomer of 60,500, with a pl of 5.5 in both strains. Purified MCE from both populations had identical K-m and V-max Values for the hydrolysis of malathion as well as for three other esterase substrates. Similarly, the kinetics of inhibition by several inhibitors were the same for the MCE from each strain. These data therefore suggest that resistance to malathion is due to a quantitative rather than a qualitative change in the MCE of the two strains. Estimation of the total MCE content in each strain showed that the resistant blowflies had nine times more MCE than the more susceptible insects. Although blowfly MCE showed greater specificity for naphthyl esters over malathion, it nevertheless hydrolyzes malathion faster than any other esterase yet isolated from an insect. This is in sharp contrast to previously studied insect strains in which organophosphate resistance has been attributed to large increases in nonspecific esterases that show very slow or no hydrolysis of the insecticides.

MEDLINE ANSWER 11 OF 13

ACCESSION NUMBER: 94304400 MEDLINE

DOCUMENT NUMBER: 94304400 PubMed ID: 8031294

TITLE: A cluster of esterase genes on chromosome 3R of Drosophila

melanogaster includes homologues of esterase genes

conferring insecticide resistance in Lucilia

cuprina.

Spackman M E; Oakeshott J G; Smyth K A; Medveczky K M; AUTHOR:

Russell R J

CSIRO Division of Entomology, Canberra, ACT. CORPORATE SOURCE:

BIOCHEMICAL GENETICS, (1994 Feb) 32 (1-2) 39-62. SOURCE:

Journal code: 0126611. ISSN: 0006-2928.

PUB. COUNTRY: United States

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals

ENTRY MONTH: 199408

Entered STN: 19940818 ENTRY DATE:

> Last Updated on STN: 19940818 Entered Medline: 19940805

We identify an esterase isozyme in Drosophila melanogaster, EST 23, which AB shares biochemical, physiological, and genetic properties with esterase E3, which is involved in resistance to organophosphate insecticides in Lucilia cuprina. Like E3, the D. melanogaster EST 23 is a membrane-bound alpha-esterase which migrates slowly toward the anode at pH 6.8. Both enzymes have similar preferences for substrates with shorter acid side chain lengths. Furthermore, on the basis of their high sensitivity to inhibition by paraoxon and their insensitivity to inhibition by eserine sulfate, both enzymes were classified as subclass I carboxylesterases. The activity of each enzyme peaks early in development and, again, in the adult stage. Both enzymes are found in the male reproductive system and larval and adult digestive tissues, the latter being consistent with a role for these enzymes in organophosphate resistance. Fine structure deficiency mapping localized Est 23 to

cytological region 84D3 to E1-2 on the right arm of chromosome 3. Moreover, we show that the genes encoding three other esterase phenotypes also map to the same region; these phenotypes involve allozymic differences in EST 9 (formerly EST C), ali-esterase activity, defined by the hydrolysis of methyl butyrate, and malathion carboxylesterase activity, defined by hydrolysis of the organophosphate malathion. This cluster corresponds closely to that encompassing E3 and malathion carboxylesterase on chromosome 4 in L. cuprina, the homologue of chromosome 3R in D. melanogaster.

ANSWER 12 OF 13 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC.DUPLICATE 1.5

ACCESSION NUMBER: 1994:272233 BIOSIS

DOCUMENT NUMBER:

PREV199497285233

TITLE:

Insecticide resistance and malathion

carboxylesterase in the sheep blowfly, Lucilia

cuprina.

AUTHOR(S):

Whyard, Steven; Russell, Robyn J.; Walker, Virginia K. (1)

(1) Dep. Biol. Insect Biotech Canada, Queen's Univ., CORPORATE SOURCE:

Kingston, ON K7L 3N6 Canada

SOURCE:

Biochemical Genetics, (1994) Vol. 32, No. 1-2, pp. 9-24.

ISSN: 0006-2928.

DOCUMENT TYPE:

Article English

LANGUAGE: Resistance to the organophosphorus insecticide malathion in genetically related strains of the Australian sheep blowfly Lucilia cuprina was examined. Separate lines of blowflies were established by homozygosis of the fourth chromosome of the parental RM strain. Both the RM and the derived resistant (der-R) strains are approximately 100 times more resistant to malathion than the related susceptible der-S strain, resistance being correlated with a 45- to 50-fold increase in a malathion carboxylesterase (MCE) activity. MCE has a pH optimum ranging between 6.6 and 8.0 and is strongly inhibited by the carboxylesterase inhibitors triphenyl phosphate, paraoxon, and diisopropylfluorophosphate. Subcellular fractionation revealed that MCE was localized predominantly to the cytosol and mitochondria in both resistant and susceptible blowflies. A single MCE was purified to homogeneity from PM blowflies. It has a pI of 5.5, is a monomer of 60.5 kDa, and hydrolyzes malathion with a V-max of 755 nmol/min/mg protein and a K-m of 11.0 mu-M. L. cuprina have thus evolved a remarkable MCE which is faster and more efficient at hydrolyzing a specific insecticide than any other insect esterase yet described.

L5 - ANSWER 13 OF 13 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC.DUPLICATE 7

ACCESSION NUMBER:

CORPORATE SOURCE:

1987:399293 BIOSIS

DOCUMENT NUMBER:

BA84:75473

TITLE:

GENERAL ESTERASE MALATHION

CARBOXYLESTERASE AND MALATHION RESISTANCE IN CULEX-

TARSALIS.

AUTHOR(S):

ZIEGLER R; WHYARD S; DOWNE A E R; WYATT G R; WALKER V K DEP. BIOCHEM., BIOSCIENCES W., UNIV. ARIZ., TUSCON, ARIZ.

85721.

SOURCE:

PESTIC BIOCHEM PHYSIOL, (1987) 28 (2), 279-285.

CODEN: PCBPBS. ISSN: 0048-3575.

FILE SEGMENT:

BA; OLD

LANGUAGE:

English

The role of esterases in malathion resistance in Culex tarsalis has been investigated. When larvae of a resistant and a sensitive strain were placed in water containing [14C] malathion, malathion penetrated to give initially similar internal levels. With resistant mosquitoes, after 15 min the internal malathion concentration decreased to low levels while the monoacid degradation products accumulated in the larvae and were

excreted into the surrounding water, whereas in susceptible larvae the internal malathion level stayed high and was lethal. It is suggested that the decrease in internal malathion and the resulting resistance were caused by an active malathion carboxylesterase in the resistant strain. A specific assay for  ${\bf malathion}$ carboxylesterase with [14C] malathion showed 55 times more activity in resistant than in susceptible larvae, whereas when general esterase activity was assayed with .alpha.-naphthyl acetate only 1.7 times the activity was found. Analyses by starch gel electrophoresis showed a peak of malathion carboxylestease, 60-fold higher from resistant than from susceptible larvae, in a gel zone which did not strain for general esterase activity. General esterases that did not hydrolyze malathion showed different electrophoretic patterns in the two populations, which are likely due to the nonisogenic character of the strains. These results show that use of a specific assay and the demonstration of degradation of malathion in vivo are essential for assessment of the contribution of esterase activity to the malathion-resistant phenotype in mosquito populations.

=> d his (FILE 'HOME' ENTERED AT 14:20:37 ON 22 MAY 2003) INDEX 'ADISCTI, ADISINSIGHT, ADISNEWS, AGRICOLA, ANABSTR, AQUASCI, BIOBUSINESS, BIOCOMMERCE, BIOSIS, BIOTECHABS, BIOTECHDS, BIOTECHNO, CABA, CANCERLIT, CAPLUS, CEABA-VTB, CEN, CIN, CONFSCI, CROPB, CROPU, DDFB, DDFU, DGENE, DRUGB, DRUGLAUNCH, DRUGMONOG2, ... ENTERED AT 14:21:05 ON 22 MAY 2003 SEA CARBOXYLESTERASE 17 FILE ADISCTI FILE ADISINSIGHT 3 FILE ADISNEWS 2 FILE AGRICOLA 335 FILE ANABSTR 33 69 FILE AQUASCI 90 FILE BIOBUSINESS 1 FILE BIOCOMMERCE FILE BIOSIS 1624 FILE BIOTECHABS 182 FILE BIOTECHDS 182 FILE BIOTECHNO 529 FILE CABA 780 244 FILE CANCERLIT FILE CAPLUS 2393

FILE CEABA-VTB

FILE CONFSCI

FILE CROPB

FILE DDFB

FILE DDFU

FILE DGENE

FILE DRUGB

FILE DRUGNL

FILE DRUGUPDATES

FILE ESBIOBASE

FILE DRUGU

FILE EMBAL FILE EMBASE

FILE FEDRIP

FILE FROSTI

FILE GENBANK FILE HEALSAFE

FILE IFIPAT

FILE KOSMET

FILE LIFESCI

FILE MEDLINE

FILE OCEAN

FILE PASCAL

FILE PHAR

FILE PHIN

FILE PROMT

FILE NIOSHTIC FILE NTIS

FILE PHARMAML

FILE SCISEARCH

FILE TOXCENTER

FILE USPATFULL

FILE USPAT2

FILE JICST-EPLUS

FILE FSTA

FILE CIN

27

1 55

104

381

134 209

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134

255

1702

474

25 11

59

14 28

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1655

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8 1332

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1229

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QUE CARBOXYLESTERASE Ll FILE 'CAPLUS, TOXCENTER, EMBASE, MEDLINE, BIOSIS, SCISEARCH' ENTERED AT 14:22:09 ON 22 MAY 2003 975 S L1 AND (ORGANOPHOSPAHTE OR MALATHION) L2 1869 S L1 AND (ORGANOPHOSPHATE OR MALATHION) L3 L4 487 S L3 AND (HYDROLYSIS OR DEGRAD?) 126 S L4 AND PY>1995 L5 51 DUP REM L5 (75 DUPLICATES REMOVED) L6

9

FILE VETB 22 FILE VETU 44 FILE WPIDS 44 FILE WPINDEX

=> d 16 ibib ab 41-51 ANSWER 41 OF 51 EMBASE COPYRIGHT 2003 ELSEVIER SCI. B.V. ACCESSION NUMBER: 97170512 EMBASE 1997170512 DOCUMENT NUMBER: Preventive effect of the extract of Du-Zhong (Tochu) leaf TITLE:

and ginseng root on acute toxicity of chlorpyrifos.

Furutsu M.; Koyama Y.-I.; Kusakabe M.; Takahashi S. AUTHOR: M. Furutsu, Department of Biochemistry, College of CORPORATE SOURCE:

Pharmacy, Nihon University, 7-7-1 Narashinodai, Funabashi,

Chiba 274, Japan

Japanese Journal of Toxicology and Environmental Health, SOURCE:

(1997) 43/2 (92-100).

Refs: 28

ISSN: 0013-273X CODEN: JJTHEC

COUNTRY: Japan

DOCUMENT TYPE: Journal; Article

Clinical Biochemistry FILE SEGMENT: 029

> 030 Pharmacology

Drug Literature Index 037

052 Toxicology

LANGUAGE: English SUMMARY LANGUAGE: English

The preventive effects of the extract (DP-extract) prepared from leaves of Eucommia ulmoides OLIVER, Eucommiaceae (Du-Zhong leaf) and roots of Panax ginseng C.A. Meyer (Ginseng), which are widely used as healthful teas, on acute toxicity of chlorpyrifos, a common organophosphorus insecticide, were investigated in mice. A 50% lethal dose of chlorpyrifos was 16.9% higher in the group pretreated with DP-extract for 3 weeks (6.66 g of dried leaves and root/kg/d, p.o.) than in the chlorpyrifos alone group. Cholinesterase (ChE) activities in serum were higher and the residual chlorpyrifos in liver was lower in mice pretreated with DP-extract than in the chlorpyrifos-alone group. Hepatic cytochrome P450 content, activities of NADPH-cytochrome c reductase and carboxylesterase (EC3.1.1.1) in livers of DP-extract pretreated mice were significantly higher than those of untreated control immediately after chlorpyrifos injection. The activities of those enzymes were also significantly higher in the DP-extract pretreated mice than the controls. Northern blot analysis of microsomes from livers of DP-extract pretreated mice revealed the increased transcription of NADPH-cytochrome c reductase and carboxylesterase. These results suggest that DP-extract increased the activities of cytochrome P450 and carboxylesterase and accelerated detoxification of chlorpyrifos to prevent the acute toxicity of the organophosphorus insecticide.

ANSWER 42 OF 51 EMBASE COPYRIGHT 2003 ELSEVIER SCI. B.V.

ACCESSION NUMBER: 97328778 EMBASE

DOCUMENT NUMBER: 1997328778

TITLE: Serum 'B' esterases as a nondestructive biomarker for

monitoring the exposure of reptiles to organophosphorus

insecticides.

Sanchez J.C.; Fossi M.C.; Focardi S. AUTHOR:

S. Focardi, Department of Environmental Biology, University CORPORATE SOURCE:

of Siena, 53100 Siena, Italy

Ecotoxicology and Environmental Safety, (1997) 38/1 SOURCE:

> (45-52). Refs: 34

ISSN: 0147-6513 CODEN: EESADV

United States COUNTRY:

DOCUMENT TYPE: Journal; Article

FILE SEGMENT: 046 Environmental Health and Pollution Control

> 052 Toxicology

LANGUAGE: English SUMMARY LANGUAGE: English

A field study was conducted to validate serum B esterases as AΒ nondestructive biomarkers (NDBs) in lizards. Serum butyrylcholinesterase (BChE) and carboxylesterase (CbE) activities were measured in lizards and four species of birds collected in an area of 0.5 ha sprayed with 0.36 kg a.i./ha of Folidol SE5 (5% parathion). Serum B esterase activities were determined in a total of 213 lizards (Gallotia galloti) and 81 birds of four species (Sylvia melanocephala, Serinus canaria, Parus caeruleus, and Eritltacus rubecula) collected for 23 days after the spraying. A control group of 39 lizards and 58 birds was sampled before the spraying. No relationship was found between serum B esterases and sex or biometric parameters in all species. Inhibition of BChE (>40%) and CbE (>50%) activities was recorded in lizards 23 days after spraying. BChE activity was found to be more sensitive than CbE to inhibition by parathion. Inhibition of serum B esterase activities was recorded in only two bird species (S. melanocephala and S. canaria), but the number of individuals collected was much less than the lizards; The advantages and disadvantages of G. galloti as bioindicator of exposure to organophosphorus insecticides in the Canary Islands (Spain) are discussed in relation to birds commonly used for this purpose.

L6 ANSWER 43 OF 51 CAPLUS COPYRIGHT 2003 ACS DUPLE

DUPLICATE 24

ACCESSION NUMBER:

1997:448523 CAPLUS

DOCUMENT NUMBER:

127:172573

TITLE:

Biochemistry of esterases associated with organophosphate resistance in Lucilia cuprina with comparisons to putative orthologs in other

Diptera

AUTHOR(S):

Campbell, Peter M.; Trott, Josephine F.; Claudianos,

Charles; Smyth, Kerrie-Ann; Russell, Robyn J.;

Oakeshott, John G.

CORPORATE SOURCE:

Div. Entomology, CSIRO, Canberra, 2601, Australia

SOURCE:

Biochemical Genetics (1997), 35(1/2), 17-40 CODEN: BIGEBA; ISSN: 0006-2928

PUBLISHER: Plenum

DOCUMENT TYPE:

Journal

LANGUAGE: English Esterase activities assocd. with organophosphate insecticide resistance in the Australian sheep blowfly, Lucilia cuprina, are compared with similar activities in other Diptera. The enzymes making the major contribution to Me butyrate hydrolysis ("ali-esterase") in L. cuprina, M. domestica, and D. melanogaster comigrate during electrophoresis. The enzymes in L. cuprina and D. melanogaster correspond to the naphthyl acetate hydrolyzing E3 and EST23 isoenzymes of those species. These and previously published data suggest that the ali-esterases of all 3 species are orthologous. Strains of L. cuprina fall into 4 groups on the basis of quant. detns. of their ali-esterase, OP hydrolase, and malathion carboxylesterase activities and these groups correspond to their status with respect to 2 types of OP resistance. Strains susceptible to OPs have high ali-esterase, low OP hydrolase, and intermediate MCE activities; those resistant to malathion but not diazinon have low ali-esterase, intermediate OP hydrolase, and high MCE activities; those resistant to diazinon but not malathion have low ali-esterase, high OP hydrolase, and low MCE activities; those resistant to both OPs have low ali-esterase, high OP hydrolase, and high MCE activities. The correlated changes among the 3 biochem. and 2 resistance phenotypes suggest that they are all properties of one gene/enzyme system; 3 major allelic variants of that system explain OP susceptibility and the 2 types of OP resistance. Models are proposed to explain the joint contribution of OP hydrolase and MCE activities to malathion resistance and the invariant assocn. of low ali-esterase and elevated OP hydrolase activities in either type of resistance.

L6 ANSWER 44 OF 51 SCISEARCH COPYRIGHT 2003 THOMSON ISI ACCESSION NUMBER: 97:31717 SCISEARCH

THE GENUINE ARTICLE: VZ611

TITLE:

Interspecies differences in enzymes reacting with organophosphates and their inhibition by paraoxon

in vitro

AUTHOR:

KalisteKorhonen E (Reprint); Tuovinen K; Hanninen O UNIV KUOPIO, NATL LAB ANIM CTR, POB 1627, FIN-70211 CORPORATE SOURCE:

KUOPIO, FINLAND (Reprint); UNIV KUOPIO, DEPT PHYSIOL,

FIN-70211 KUOPIO, FINLAND

COUNTRY OF AUTHOR:

FINLAND

SOURCE:

HUMAN & EXPERIMENTAL TOXICOLOGY, (DEC 1996) Vol.

15, No. 12, pp. 972-978.

Publisher: STOCKTON PRESS, HOUNDMILLS, BASINGSTOKE,

HAMPSHIRE, ENGLAND RG21 6XS.

ISSN: 0144-5952.

DOCUMENT TYPE: FILE SEGMENT:

Article; Journal

LANGUAGE:

LIFE English

REFERENCE COUNT:

23

\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

1 Inhibition of cholinesterases (ChE) and carboxylesterases (CaE) by paraoxon (Px) was studied in vivo in the serum, liver, lung and muscle of mouse, guineapig, rabbit and man (serum only). Moreover, the role of Px hydrolyzing enzyme (Pxase) in the detoxification of Px was studied by inhibiting its activity with EDTA.

2 The ChE and CaE activities as well as their sensitivity to Px varied in different tissues and species. The ChEs were more sensitive than CaEs to Px except in the liver. The CaE activity in human and rabbit sera was low and resistant to Px, indicating that it may have a minor importance

for the binding of Px. 3 The Px-inhibited ChEs were spontaneously reactivated in the mouse and rabbit sera during 24 h. In mouse, also the CaE activity was recovered. The presence of EDTA in the incubation medium prevented this reactivation indicating that Pxase takes part in the reactivation process.

4 In rabbit, the serum Pxase activity was very high suggesting a good

Px detoxifying capacity of the rabbit serum.

5 The results show that amounts and sensitivities of esterases to OPs in rodents may markedly differ from that in man. Possible species-related differences in the affinity of ChEs and CaEs for OPs and the OP hydrolyzing activity should be taken into the consideration, when animal data are extrapolated to man.

ANSWER 45 OF 51 CAPLUS COPYRIGHT 2003 ACS DUPLICATE 25

ACCESSION NUMBER:

1996:743517 CAPLUS

DOCUMENT NUMBER:

126:140649

TITLE:

Hydrolysis of parabenes by extracts from

differing layers of human skin

AUTHOR(S):

Lobemeier, Claudia; Tschoetschel, Carla; Westie,

Sonja; Heymann, Eberhard

CORPORATE SOURCE:

Physiological Chem., Univ. Osnabrueck, Osnabrueck,

D-49069, Germany

SOURCE:

Biological Chemistry (1996), 377(10),

647-651

CODEN: BICHF3; ISSN: 1431-6730

de Gruyter PUBLISHER: DOCUMENT TYPE: Journal LANGUAGE: English

Four carboxylesterases were capable of hydrolyzing

4-hydroxybenzoic acid esters in human skin and s.c. fat tissue. The highest specific activities were found in an ext. from s.c. fat tissue. The most prominent esterase of this tissue prefers the Me ester of 4-hydroxybenzoic ester (Me parabene). Its activity decreases with increasing chain length of the alc. moiety of the parabenes. The existence of a 2nd parabene esterase in s.c. fat is concluded from organophosphate inhibition characteristics. Another prominent

parabene esterase was characterized in exts. from transformed keratinocytes. It prefers Bu parabene and its activity decreases with decreasing chain length of the alc. moiety. The 4th parabene esterase is an enzyme in blood which contaminates the tissue exts. used here. All of the tissue exts. were active at pH 8.0.

ANSWER 46 OF 51 SCISEARCH COPYRIGHT 2003 THOMSON ISI

ACCESSION NUMBER:

96:338761 SCISEARCH

THE GENUINE ARTICLE: UG570

TITLE:

INTERACTION OF ORGANOPHOSPHORUS COMPOUNDS WITH

CARBOXYLESTERASES IN THE RAT

AUTHOR:

JOKANOVIC M (Reprint); KOSANOVIC M; MAKSIMOVIC M

CORPORATE SOURCE:

UNIV PADUA, IST MED LAVORO, VIA FACCIOLATI 71, I-35127 PADUA, ITALY (Reprint); FAC PHARM BELGRADE, DEPT TOXICOL,

YU-11000 BELGRADE, YUGOSLAVIA

COUNTRY OF AUTHOR:

ITALY; YUGOSLAVIA

SOURCE:

ARCHIVES OF TOXICOLOGY, (APR 1996) Vol. 70, No.

7, pp. 444-450. ISSN: 0340-5761.

DOCUMENT TYPE:

Article; Journal LIFE

FILE SEGMENT: LANGUAGE:

ENGLISH

REFERENCE COUNT:

\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

Carboxylesterases (CarbE) are involved in detoxication of AB organophosphorus compounds (OPC) through two mechanisms: hydrolysis of ester bonds in OPC which contain them and binding of OPC at the active site of CarbE which reduces the amount of OPC available for acetylcholinesterase inhibition. This study of the interaction of rat plasma and liver CarbE with dichlorvos, soman and sarin in vitro and in vivo was undertaken in order to contribute to better understanding of the role of CarbE in detoxication of OPC. The results obtained have shown that inhibitory potency (I-50) of dichlorvos, sarin and soman towards rat liver CarbE was 0.2 mu M, 0.5 mu M and 4.5 mu M, respectively, for 20-min incubation at 25 degrees C. Second-order rate constants (k(a)) for liver CarbE inhibition were 2.3 x 10(5) M(-1) min(-1), 6.9 x 10(4) M(-1) min(-1) and  $1.1 \times 10(4) \text{ M}(-1) \text{ min}(-1)$  for dichlorvos, sarin and soman, respectively. The corresponding values for plasma CarbE could not be calculated because of dominant spontaneous reactivation of inhibited CarbE. CarbE inhibited with these OPC in vitro spontaneously reactivate with half-times of 18, 143 and 497 min for sarin, dichlorvos and soman in plasma and 111, 163 and 297 mill for sarin, soman and dichlorvos in liver, respectively. These results were also confirmed in experiments in vivo in which rats were subcutaneously treated with 0.5 LD(50) of these agents. The half-times of spontaneous reactivation of rat plasma CarbE in vivo were 1.2, 2.0 and 2.7 h for dichlorvos, sarin and soman, respectively. These findings have changed current understanding of the mechanism of interaction of CarbE with OPC and involvement of the enzymes in detoxication of OPC, suggesting an active and important role of the

enzymes in metabolic conversions of OPC to their less toxic metabolites.

ANSWER 47 OF 51 SCISEARCH COPYRIGHT 2003 THOMSON ISI

ACCESSION NUMBER:

96:785530 SCISEARCH

THE GENUINE ARTICLE: VN707

TITLE:

EPTASTIGMINE-PHOSPHOTRIESTERASE COMBINATION IN DFP

INTOXICATION

AUTHOR:

TUOVINEN K (Reprint); KALISTEKORHONEN E; RAUSHEL F M;

HANNINEN O

CORPORATE SOURCE:

UNIV KUOPIO, DEPT PHYSIOL, POB 1627, SF-70211 KUOPIO, FINLAND (Reprint); UNIV KUOPIO, NATL LAB ANIM CTR,

SF-70211 KUOPIO, FINLAND; TEXAS A&M UNIV, DEPT CHEM,

COLLEGE STN, TX, 77843

COUNTRY OF AUTHOR:

FINLAND; USA

SOURCE:

TOXICOLOGY AND APPLIED PHARMACOLOGY, (OCT 1996)

Vol. 140, No. 2, pp. 364-369.

ISSN: 0041-008X. Article; Journal

FILE SEGMENT: LANGUAGE:

LIFE ENGLISH

REFERENCE COUNT:

DOCUMENT TYPE:

\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

A novel therapy against organophosphate exposure, the AΒ combination of a carbamate eptastigmine and an organophosphate hydrolase (phosphotriesterase) was studied in mice against diisopropylfluorophosphate (DFP) (1.75 mg/kg) exposure. Mice received eptastigmine (0.9 mg/kg; iv) 10 min prior to the ip injection of DFP. Phosphotriesterase (83 U/g body weight) was injected iv 10 min after DFP. Eptastigmine (1.5 mg/kg; iv) inhibited the acetylcholinesterase activities in brain and erythrocytes for a longer time than physostigmine, Eptastigmine caused only minor changes in the behavior and activity of the animals, whereas physostigmine clearly reduced their activity for about 30 min. The eptastigmine pretreatment clearly supplemented the protective effect of phosphotriesterase against DFP: the plasma butyrylcholinesterase activity was doubled and the activity recovered faster than in animals treated with phosphotriesterase alone. In lung, butyrylcholinesterase activity was initially lower after eptastigmine- phosphotriesterase than phosphotriesterase treatment alone, However, the activity returned 24 hr later to normal in eptastigmine-phosphotriesterase-treated groups, With phosphotriesterase only, it recovered only to 75% of the control level. Presumably eptastigmine, by preventing the binding of DFP to cholinesterases, caused an elevation of free DFP levels in body fluids and promoted phosphotriesterase hydrolysis of DFP. (C) 1996 Academic Press, Inc.

ANSWER 48 OF 51 CAPLUS COPYRIGHT 2003 ACS

DUPLICATE 26

ACCESSION NUMBER: 1996:292073 CAPLUS

DOCUMENT NUMBER:

124:335641

TITLE:

Inhibition by insecticides of partially purified

carboxylesterase from Aphis gossypii

(Homoptera: Aphididae)

AUTHOR(S):

CORPORATE SOURCE:

Owusu, Ebenezer O.; Horiike, Michio; Hirano, Chisato Pesticide Research Laboratory, Kochi University,

Nankoku, 783, Japan

SOURCE .

Journal of Economic Entomology (1996),

89(2), 307-310

CODEN: JEENAI; ISSN: 0022-0493 Entomological Society of America

DOCUMENT TYPE:

Journal

PUBLISHER: LANGUAGE:

English

Inhibition of partially purified carboxylesterase from a dichlorvos selected (E-D-R) strain of cotton aphid, Aphis gossypii (Glover), by selected insecticides was demonstrated in vitro. Regeneration of active enzymes was not obsd. up to 5 h after removal of excess inhibitor. This result indicates that pesticide hydrolysis is not a likely resistance mechanism in A. gossypii. Carboxylesterases from this aphid seem likely to bind permanently to inhibitors, rendering them ineffective and thus protecting the active sites of pesticide inhibition. Of 2 organophosphate and 2 carbamate insecticides tested, dichlorvos was most inhibitory. The order of toxicity ranked dichlorvos > naled > carbaryl > m-tolyl methylcarbamate.

ANSWER 49 OF 51 SCISEARCH COPYRIGHT 2003 THOMSON ISI

ACCESSION NUMBER: 96:227285 SCISEARCH

THE GENUINE ARTICLE: UA518

TITLE:

THE PERSISTENCE AND FATE OF MALATHION RESIDUES

IN STORED BEANS (PHASEOLUS-VULGARIS) AND MAIZE (ZEA-MAYS)

LALAH J O; WANDIGA S O (Reprint) AUTHOR:

UNIV NAIROBI, COLL BIOL & PHYS SCI, DEPT CHEM, POB 30197, CORPORATE SOURCE:

NAIROBI, KENYA (Reprint); UNIV NAIROBI, COLL BIOL & PHYS

SCI, DEPT CHEM, NAIROBI, KENYA

COUNTRY OF AUTHOR:

KENYA

SOURCE:

PESTICIDE SCIENCE, (MAR 1996) Vol. 46, No. 3,

pp. 215-220. ISSN: 0031-613X.

DOCUMENT TYPE:

Article; Journal

FILE SEGMENT: LANGUAGE:

AGRI ENGLISH

REFERENCE COUNT:

\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

Two experimental models simulating the traditional storage conditions AB prevalent in Kenya, i.e. the open basket model and the modern wooden box model, were used to study the rate of dissipation and fate of malathion residues in maize grains and beans stored for periods of up to one year at ambient temperatures averaging 23 degrees C. The grain samples were initially treated with 10.36 mg kg(-1) of radiolabelled malathion dust prior to storage and portions analysed at regular intervals for malathion, malaoxon and the transformation products isomalathion, malathion alpha-monocarboxylic acid and malathion beta-monocarboxylic acid using a combination of chromatographic, radioisotopic and mass-spectrometric techniques.

The findings showed a gradual penetration of malathion into the grains in amounts which were slightly higher in maize than in beans irrespective of the method of storage. After 51 weeks of storage, 34-60% of the initial residues persisted in all the grains. The total residual levels were slightly higher in beans than in maize irrespective of the storage methods though the persistence was a little higher in the wooden box than in the open basket. The rates of dissipation of the pesticide from the grains decreased with storage time and followed a biphasic pattern. Applying first-order reaction kinetics, the following half-lives were obtained: maize grains stored in open basket: 194 days; maize grains stored in closed wooden box: 261 days; beans stored in open basket: 259 days; beans stored in closed wooden box: 405 days. Beans stored in the wooden box had higher levels of bound residues than those sampled from the open basket. This trend was similar in maize grains although the concentrations were lower. The analysis of malathion metabolites confirmed the degradation trend of the residues.

ANSWER 50 OF 51 CAPLUS COPYRIGHT 2003 ACS

ACCESSION NUMBER: 1996:358942 CAPLUS

DOCUMENT NUMBER:

125:28019

TITLE:

Protection of organophosphate-inactivated

esterases with phosphotriesterase

AUTHOR(S):

Tuovinen, Kai; Kaliste-Korhonen, Eila; Raushel, Frank

M.; Hanninen, Osmo

CORPORATE SOURCE:

Dep. Physiology, Univ. Kuopio, Kuopio, FIN-70211,

Finland

SOURCE:

Fundamental and Applied Toxicology (1996),

31(2), 210-217

CODEN: FAATDF; ISSN: 0272-0590

PUBLISHER: DOCUMENT TYPE: Academic

Journal English LANGUAGE:

The protective effect of phosphotriesterase (PTE) on cholinesterase (ChE) AB and carboxylesterase (CaE) activities was studied in mice. The PTE per-treatment (120 U/g body wt., 9.6 .mu.g/g body wt.) given i.v. 10 min before diisopropyl fluorophosphate, sarin, or soman variably prevented ChE inhibition in erythrocytes and plasma and CaE in plasma. PTE also protected the brain and lung ChEs against inactivation by organophosphates (OPs). The recovery of the enzymes was dependent on the OP used. Postexposure therapy with PTE, given 1.5 h after paraoxon, also prevented ChE inhibition in erythrocytes, brain, and lung

24 h after exposure. The distribution studies with [1251] PTE showed that PTE does not markedly gain access into the central nervous system.

ANSWER 51 OF 51 CAPLUS COPYRIGHT 2003 ACS

DUPLICATE 27

ACCESSION NUMBER: DOCUMENT NUMBER:

1996:352828 CAPLUS

125:28249

TITLE:

Purification and characterization of

carboxylesterases of a rice green leafhopper

Nephotettix cincticeps

AUTHOR(S):

Chiang, Shih-Wen; Sun, Chih-Ning

CORPORATE SOURCE:

Dep. Entomology, National Chung-Hsing Univ., Taichung,

40227, Taiwan

SOURCE:

Pesticide Biochemistry and Physiology (1996

), 54(3), 181-189

CODEN: PCBPBS; ISSN: 0048-3575

PUBLISHER: DOCUMENT TYPE: Academic Journal

English LANGUAGE: More than four carboxylesterase isoenzymes in the homogenate of

susceptible strain.

a rice green leafhopper could be resolved by isoelec. focusing electrophoresis. A combination of ammonium sulfate fractionation, gel filtration, and chromatofocusing chromatog. was used to isolate and purify these isoenzymes. Four fractions, i.e., E1, E2, E3, and E4, with pI's ranging from 5.1 to 4.85, were obtained. The most abundant E3 had a mol. mass of 58.6 kDa and appeared electrophoretically homogeneous on SDS-PAGE. [1,3-3H]Diisopropyl fluorophosphate-labeling expt. revealed that the proteins of 58.6 kDa, a minor component of E2 and the major component of E4, were the carboxylesterase isoenzymes sought. A protein of the same mol. wt. which existed in a very minute amt. in El and was barely detectable on SDS-PAGE by Coomassie blue staining was actually the carboxylesterase isoenzyme of pI 5.1. All four fractions exhibited significant activity toward several mode substrates with .alpha.-naphthyl butyrate being the most preferred. Their activity toward malathion, permethrin, and cypermethrin was ca. 106-fold lower than their activity toward the model substrates. The pyrethroids were hydrolyzed more readily than malathion by these hydrolases, and cis-permethrin was more preferred than the trans-isomer. E4 was the only fraction that cross-reacted with the antiserum against carboxylesterases of a rice brown planthopper, Nilapavata lugens. Among the four isoenzyme fractions, E3, the most abundant, showed low activity toward all four insecticides and was the least active fraction toward cis-permethrin and cypermethrin. A field strain of N. cincticeps had 26- to 37-fold higher carboxylesterase activity toward the model substrates than a susceptible strain. Yet, little, if any, difference in the hydrolysis of malathion, permethrin, and cypermethrin was obsd. between these two stains. The field strain produced .gtoreq.8 times more carboxylesterases than the

ANSWER 30 OF 51 CAPLUS COPYRIGHT 2003 ACS 2000:56242 CAPLUS ACCESSION NUMBER:

DOCUMENT NUMBER:

132:344746

TITLE:

SOURCE:

In vitro estimation of the enzymatic

hydrolysis of malathion and its

O, O-dialkyl analogs (C1-C4) enantiomers

Polec, Iwona; Legocki, Jan; Czajka, Magdalena

AUTHOR(S): Instytut Przemyslu Organicznego, Warsaw, 03-236, Pol. CORPORATE SOURCE:

Organika (1999), Volume Date 1997-1998, (Pt.

2), 21-30

CODEN: ORGAD2; ISSN: 0137-9933 Instytut Przemyslu Organicznego

PUBLISHER: DOCUMENT TYPE:

Journal

LANGUAGE:

Polish

Malathion [S-1,2-di(ethoxycarbonyl)ethyl-0,0-dimethyl

dithiophosphate], and its 0,0-dialkyl derivs. (R = C1-C4) enantiomers were

exposed to the action of carboxylesterase from rabbit liver.

Michaelis consts. (Km) and maximal enzymic reaction rates (Vmax) were detd. for each of examd. compds. HPLC was used as the anal. method; the quantity of studied substrate was measured after the limited unit of time of the enzymic hydrolysis duration, and then compared with the adequate ref. pattern. It has been concluded that in the range of used substrates concns., the affinity of enzyme (expressed as Km) increased with an increase of the no. of carbon atoms in alkyl chain. Maximum rate of enzymic hydrolysis has been shown for O, O-di-Et

malathion deriv. No clear relationship between estd. Km and Vmax values, and configuration of the tested compds. enantiomers was found.

ANSWER 31 OF 51 SCISEARCH COPYRIGHT 2003 THOMSON ISI

ACCESSION NUMBER:

1999:36043 SCISEARCH

THE GENUINE ARTICLE: 151TY

TITLE:

Chicken serum albumin hydrolyzes dichlorophenyl phosphoramidates by a mechanism based on transient

phosphorylation

AUTHOR:

Sogorb M A; Monroy A; Vilanova E (Reprint)

CORPORATE SOURCE:

UNIV MIGUEL HERNANDEZ, FAC MED, UPD GENET NUTR & TOXICOL, DIV TOXICOL, E-03550 ALICANTE, SPAIN (Reprint); UNIV

MIGUEL HERNANDEZ, INST BIOINGN, UNIDAD TOXICOL & SEGURIDAD

QUIM, ALICANTE, SPAIN

COUNTRY OF AUTHOR:

SOURCE:

CHEMICAL RESEARCH IN TOXICOLOGY, (DEC 1998) Vol.

11, No. 12, pp. 1441-1446.

Publisher: AMER CHEMICAL SOC, 1155 16TH ST, NW,

WASHINGTON, DC 20036.

ISSN: 0893-228X.

DOCUMENT TYPE: FILE SEGMENT:

Article; Journal

LANGUAGE:

English

LIFE

SPAIN

REFERENCE COUNT: 3 1

\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

The hydrolyzing activities of O-hexyl O-2,5-dichlorophenyl AB phosphoramidate (HDCP, and p-nitrophenyl butyrate (p-NPB) in chicken serum had been found to copurify in the same protein, identified as albumin, The hydrolyzing activities of both chicken ser um and commercial serum albumins from different species were inhibited in a dose-dependent manner by short chain fatty acids. On simultaneous incubation of chicken serum with HDCP and p-NPB, a competitive interaction was detected between the two substrates, This behavior suggests that both are hydrolyzed in the same albumin active site. When chicken serum was preincubated with one of the substrates, and the latter were withdrawn by large dilution, the hydrolyzing activities with both substrates were found to be reduced. This reduction was in turn dependent upon the time of preincubation with the first substrate. These results suggest that HDCP and p-NPB are hydrolyzed

by the same albumin active site, via a mechanism based on transient phosphorylation/acylation of the active site. The proposed **hydrolysis** mechanism would account for the hydrolytic kinetics of both substrates.

L6 ANSWER 32 OF 51 SCISEARCH COPYRIGHT 2003 THOMSON ISI

ACCESSION NUMBER: 1998:362837 SCISEARCH

THE GENUINE ARTICLE: ZL902

TITLE:

Cross-resistance patterns among Lucilia cuprina (Diptera: Calliphoridae) resistant to organophosphorus insecticides Campbell P M (Reprint); Yen J L; Masoumi A; Russell R J;

AUTHOR: Campbell P M (Reprint); Yen J L; Masoumi Batterham P; McKenzie J A; Oakeshott J G

CORPORATE SOURCE: CSIRO, DIV ENTOMOL, POB 1700, CANBERRA, ACT 2601,

AUSTRALIA (Reprint)

COUNTRY OF AUTHOR:

SOURCE:

JOURNAL OF ECONOMIC ENTOMOLOGY, (APR 1998) Vol.

91, No. 2, pp. 367-375.

Publisher: ENTOMOL SOC AMER, 9301 ANNAPOLIS RD, LANHAM, MD

20706.

AUSTRALIA

ISSN: 0022-0493. Article; Journal

FILE SEGMENT: LANGUAGE:

DOCUMENT TYPE:

AGRI English

REFERENCE COUNT:

43
\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

Strains of Lucilia cuprina (Wiedemann) have been characterized as AB having low, intermediate, or high levels of esterase-mediated hydrolysis Of the organophosphorus insecticide, chlorfenvinphos. These levels correlate respectively with susceptibility to organophosphorus insecticides, malathion resistance, or diazinon resistance. Diazinon and chlorfenvinphos are diethyl organophosphorus insecticides having 2 ethoxy groups attached to their central phosphorus atom, whereas malathion is a dimethyl organophosphorus insecticide having 2 methoxy groups attached to its phosphorus atom, and, unusually, malathion also has 2 carboxylester bonds in addition to the phosphoester bonds that define organophosphorus compounds. We tested larvae for resistance to diazinon and also assessed representative malathion-resistant and diazinon-resistant L. cuprina strains at the adult stage for resistance to 12 organophosphorus insecticides, including analog pairs differing only in respect to their dimethyl-diethyl status. Two malathion-resistant strains have low-level cross-resistance to diazinon (3 to 4-fold), 4 diazinon-resistant strains have high-level diazinon resistance (11 to 16-fold), and 2 strains with a combined (malathion plus diazinon) resistance type also have high-level diazinon resistance (17 to 18-fold) relative to 3 organophosphorus insecticide-susceptible strains. One of the diazinon-resistant strains showed approximate to 2 times greater resistance factors toward diethyl organophosphorus insecticides than their dimethyl analogs while (leaving aside malathion to consider only the majority which have no carboxylester groups) a malathion -resistant strain showed 2-5 times greater resistance factors toward the dimethyl organophosphorus insectides than their diethyl analogs. The diazinon-resistant strain showed no resistance to 2 di-isopropyl organophosphorus compounds or to 2 organophosphorus insecticides which are asymmetric about the phosphorus atom (optically active). The malathion-resistant strain showed only slight resistance (<3-fold) to either the di-isopropyl or optically active organophosphorus insecticides, including the di-isobropyl analog of malathion. These cross-resistance patterns parallel those of certain organophosphorus insecticide-resistant strains of Musca domestica L., in which diazinon and malathion resistances also are proposed to be esterase mediated, reinforcing other biochemical data suggesting a general mechanism among the higher Diptera.

ANSWER 33 OF 51 CAPLUS COPYRIGHT 2003 ACS DUPLICATE 20

ACCESSION NUMBER: 1998:291581 CAPLUS 129:50806 DOCUMENT NUMBER:

Malathion-specific resistance in a strain of TITLE:

the rust red grain beetle Cryptolestes ferrugineus

(Coleoptera: Cucujidae)

Spencer, A. G.; Price, N. R.; Callaghan, A. AUTHOR(S):

School of Animal and Microbial Sciences, University of CORPORATE SOURCE:

Reading, Reading, RG6 6AJ, UK

Bulletin of Entomological Research (1998), SOURCE:

88(2), 199-206

CODEN: BEREA2; ISSN: 0007-4853

CAB International PUBLISHER:

Journal DOCUMENT TYPE: English LANGUAGE:

A strain of Cryptolestes ferrugineus (Stephens) bred for malathion -specific resistance was found to be 650 fold resistant at LD50 when compared with a susceptible strain bred from the same stock. Resistance was >98% synergized by tri-Ph phosphate and S,S,S-tri-Bu phosphorotrithioate, but unaffected by piperonyl butoxide. AChE inhibition by malaoxon varied slightly between the strains. Non-specific esterase activity as measured by the hydrolysis of .alpha.-naphthyl acetate was slightly reduced in the resistant strain whereas there were no inter-strain differences in the hydrolysis of .beta.-naphthyl acetate. Products of in vitro metab. of malathion were identified by TLC and gas chromatog.-mass spectrometry as .alpha. - and .beta. -malathion mono-acids. resistance was due to the hydrolytic breakdown of malathion by a malathion-specific carboxylesterase. The rate of in vitro malathion hydrolysis was found to be 31 times greater in the resistant strain. In vitro inhibition studies indicated that resistance is attributable to a carboxylesterase unique to the resistant strain. The implications of these results are discussed in relation to work recently carried out on malathion-specific

resistance in dipterous species. THERE ARE 54 CITED REFERENCES AVAILABLE FOR THIS REFERENCE COUNT: 54

ANSWER 34 OF 51 CAPLUS COPYRIGHT 2003 ACS DUPLICATE 21

ACCESSION NUMBER: 1998:447815 CAPLUS

129:172411 DOCUMENT NUMBER:

Two different amino acid substitutions in the TITLE:

ali-esterase, E3, confer alternative types of organophosphorus insecticide resistance in the sheep

RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

blowfly, Lucilia cuprina

Campbell, Peter M.; Newcomb, Richard D.; Russell, AUTHOR(S):

Robyn J.; Oakeshott, John G.

Division of Entomology, Industrial Research CORPORATE SOURCE:

> Organisation, Cranberra, 2601, Australia Insect Biochemistry and Molecular Biology (

**1998**), 28(3), 139-150

CODEN: IBMBES; ISSN: 0965-1748

Elsevier Science Ltd. PUBLISHER:

DOCUMENT TYPE: Journal English LANGUAGE:

SOURCE:

Two types of organophosphorus (OP) insecticide resistance are assocd. with AB reduced 'ali-esterase' (E3 isoenzyme) activity in Lucilia cuprina. The 'diazinon' resistance type shows generally greater resistance for di-Et than di-Me OPs but no resistance to malathion. The ' malathion' resistance type shows generally greater resistance for di-Me than di-Et OPs, low level diazinon resistance, but exceptionally high malathion resistance (600 x susceptible), the last being attributed to hydrolysis of the carboxylester groups which are peculiar to malathion (malathion

carboxylesterase, MCE). E3 variants from diazinon resistant strains have previously been shown to have a Gly137 .fwdarw. Asp substitution that structural modeling predicts is only about 4.6 .ANG. from the .gamma. oxygen of the catalytic serine residue. Here we show that E3 variants from malathion resistant strains have a Trp251 .fwdarw. Leu substitution predicted to be about 4.3 .ANG. from that serine. We have expressed alleles of the gene encoding both resistance variants of E3 and an OP susceptible variant in a baculovirus system and compared the kinetics of their products. We find that both resistance substitutions reduce ali-esterase activity and enhance OP hydrolase activity. Furthermore the Gly137 .fwdarw. Asp substitution enhances OP hydrolase activity for a di-Et OP substrate (chlorfenvinphos) more than does the Trp251 .fwdarw. Leu substitution, which is consistent with the OP cross-resistance patterns. Trp251 .fwdarw. Leu also reduces the Km for carboxylester hydrolysis of malathion about 10-fold to 21 .mu.M, which is consistent with increased MCE activity in malathion resistant strains. We then present a model in which the malathion carboxylesterase activity of the E3-Leu251 enzyme is enhanced in vivo by its OP hydrolase activity. The latter activity enables it to reactivate after phosphorylation by malaoxon, the activated form of malathion, accounting for the exceptionally high level of resistance to malathion. We conclude that the two types of resistance can be explained by kinetic changes caused by the two allelic substitutions in the E3 enzyme.

REFERENCE COUNT:

THERE ARE 33 CITED REFERENCES AVAILABLE FOR THIS 33 RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

ANSWER 35 OF 51 EMBASE COPYRIGHT 2003 ELSEVIER SCI. B.V.

ACCESSION NUMBER:

1998200920 EMBASE

TITLE:

Purification, molecular characterization and catalytic properties of a Pseudomonas fluorescens enzyme having

cholinesterase-like activity.

AUTHOR:

Rochu D.; Rothlisberger C.; Taupin C.; Renault F.; Gagnon

J.; Masson P.

CORPORATE SOURCE:

P. Masson, Ctr. Recherches Serv. Sante Armees, Unite d'Enzymologie, BP 87, 38702 La Tronche Cedex, France.

100335.404@compuserve.com

SOURCE:

Biochimica et Biophysica Acta - Protein Structure and

Molecular Enzymology, (1998) 1385/1 (126-138).

Refs: 64

ISSN: 0167-4838 CODEN: BBAEDZ

PUBLISHER IDENT.: S 0167-4838(98)00042-9

COUNTRY:

Netherlands

DOCUMENT TYPE: FILE SEGMENT:

Journal; Article 004 Microbiology

LANGUAGE:

English

SUMMARY LANGUAGE: English An enzyme with a cholinesterase (ChE) activity, produced by Pseudomonas fluorescens, was purified to homogeneity in a three-step procedure. Analysis by non-denaturing and SDS-PAGE, and by isoelectric focusing, indicated that the enzyme was a monomer of 43 kDa, with a pI of 6.1. The N-terminal sequence, AEPLKAVGAGEGQLDIVAWPGYIEA, showed some similarities with proteins of the ChE family and a strong similarity with a protein from Escherichia coli with unknown structure and function. Cholinesterase activity at pH 7.0 and 25.degree.C was maximum with propionylthiocholine as substrate (k(cat,app)=670 min-1), followed by acetylthiocholine, and significantly lower with butyrylthiocholine. Catalytic specificity (k(cat)/K(m)) was the same for propionylthiocholine and acetylthiocholine, but was two orders of magnitude lower for butyrylthiocholine. Kinetics of thiocholine ester hydrolysis showed inhibition by excess substrate which was ascribed to binding of a second substrate molecule, leading to non-productive ternary complex (K(m)=35 .mu.M, K(SS)=0.49 mM)with propionylthiocholine). There was low or no reactivity with organophosphates and carbamates. The enzyme inhibited by

echothiophate (k(II)=0.44x102 M-1 min-1) was not reactivated by pralidoxime methiodide. However, the P. fluorescens enzyme had affinity for procainamide and decamethonium, two reversible ChE inhibitors used as affinity chromatography ligand and eluant, respectively. Although similarity of the N-terminal amino acid sequence of the enzyme with an internal sequence of ChEs is weak, its catalytic activity towards thiocholine esters, and its affinity for positively charged ligands supports the contention that this enzyme may belong to the ChE family. However, we cannot rule out that the enzyme belongs to another structural family of proteins having cholinesterase-like properties. The reaction of the enzyme with organophosphates suggests that it is a serine esterase, and currently this enzyme may be termed as having a cholinesterase-like activity. Copyright (C) 1998 Elsevier Science B.V.

ANSWER 36 OF 51 CAPLUS COPYRIGHT 2003 ACS

ACCESSION NUMBER:

1998:140667 CAPLUS

TITLE:

A physiologically based pharmacokinetic and

pharmacodynamic model for paraoxon in rainbow trout.

AUTHOR (S):

Abbas, Richat

CORPORATE SOURCE:

Otsuka America Pharmaceutical, Inc., Rockville, MD,

20850, USA

SOURCE:

Book of Abstracts, 215th ACS National Meeting, Dallas,

March 29-April 2 (1998), AGRO-103. American

Chemical Society: Washington, D. C.

CODEN: 65QTAA

DOCUMENT TYPE:

Conference; Meeting Abstract

LANGUAGE:

English

The acute toxicity of parathion is mainly due to the inhibition of acetylcholinesterase (AChE) by its active metabolite paraoxon. To quant. characterize the relationships among organophosphate insecticide exposure, target organ concn., AChE inhibition and carboxylesterase detoxification a PBPK/PD model was developed. The model structure consisted of brain, heart, liver, kidney and remainder of the body, which were interconnected by blood circulation. Exptl. detd. tissue:blood partition coeffs., synthesis and degrdn. rates of AChE, and uptake and depuration clearances of paraoxon were used for the model development. The agreement between simulated and obsd. values indicated that this model was an appropriate tool to predict organophosphate insecticide exposure, tissue distribution and the resulting toxic effect, and to quant. est. the degree of protection that carboxylesterase provided the fish when they were exposed to paraoxon.

ANSWER 37 OF 51 CAPLUS COPYRIGHT 2003 ACS

ACCESSION NUMBER:

1997:450131 CAPLUS

DOCUMENT NUMBER:

127:77923

TITLE:

Malathion carboxylesterases of

resistant Lucilia cuprina for bioremediation of

insecticide contamination

INVENTOR(S):

Russell, Robyn Joyce; Newcomb, Richard David; Campbell, Peter Malcolm; Robin, Geoffrey Charles De

Quetteville; Claudianos, Charles; Smyth, Kerrie-ann; Boyce, Thomas Mark; Oakeshott, John Graham; Brownlie,

Jeremy Colin; et al.

PATENT ASSIGNEE(S):

Commonwealth Scientific and Industrial Research Organisation, Australia; Russell, Robyn Joyce; Newcomb, Richard David; Campbell, Peter Malcolm

PCT Int. Appl., 56 pp. SOURCE:

CODEN: PIXXD2

DOCUMENT TYPE:

Patent

LANGUAGE:

English

FAMILY ACC. NUM. COUNT:

PATENT INFORMATION:

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APPLICATION NO. DATE
                          KIND DATE
      PATENT NO.
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                                                        _____
                                                                              19961122 <--
                            Al 19970529
                                                       WO 1996-AU746
      WO 9719176
           W: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE,
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PRIORITY APPLN. INFO.:
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Genes and cDNAs encoding malathion carboxylesterases of the sheep blowfly (Lucilia cuprina) that are capable of hydrolyzing at least one organophosphate selected from the group consisting of carboxylester organophosphates and dimethyloxon organophosphates are described. Genes encoding several isoenzymes are identified and the enzymes characterized and the preferred enzymes are variants of the isoenzyme encoded by the Lc.alpha.E7 gene. The preferred analogs have an amino acid substitution of Trp-251 selected from the group consisting of Leu, Ser, Ala, Ile, Val, Thr, Cys, Met and Gly. The preferred substituents are Leu and Ser. These substitutions were identified by sequencing of a no. of cloned genes from Lucilia and the orthologous enzyme from Musca domestica.

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L6 ANSWER 38 OF 51 CAPLUS COPYRIGHT 2003 ACS DUPLICATE 22 ACCESSION NUMBER: 1997:457602 CAPLUS DOCUMENT NUMBER: 127:146108
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TITLE: A single amino acid substitution converts a

carboxylesterase to an organophosphorus

hydrolase and confers insecticide resistance on a

blowfly

AUTHOR(S): Newcomb, R. D.; Campbell, P. M.; Ollis, D. L.; Cheah,

E.; Russell, R. J.; Oakeshott, J. G.

CORPORATE SOURCE: Division Entomology, Commonwealth Scientific

Industrial Research Organization, Canberra, 2601,

Australia

SOURCE: Proceedings of the National Academy of Sciences of the

United States of America (1997), 94(14),

7464-7468

CODEN: PNASA6; ISSN: 0027-8424 National Academy of Sciences

PUBLISHER: Nationa DOCUMENT TYPE: Journal

DOCUMENT TYPE: Journal LANGUAGE: English

Resistance to organophosphorus (OP) insecticides is assocd. with decreased carboxylesterase activity in several insect species. The resistance may be the result of a mutation in a carboxylesterase that simultaneously reduces its carboxylesterase activity and confers an OP hydrolase activity (the "mutant aliesterase hypothesis"). In the sheep blowfly, Lucilia cuprina, the assocn. is due to a change in a specific esterase isoenzyme, E3, which, in resistant flies, has a null phenotype on gels stained using std. carboxylesterase substrates. The authors show that an OP-resistant allele of the gene that

encodes E3 differs at 5 amino acid replacement sites from a previously described OP-susceptible allele. Knowledge of the structure of a related enzyme (acetylcholinesterase) suggests that one of these substitutions (Gly137.fwdarw.Asp) lies within the active site of the enzyme. The occurrence of this substitution is completely correlated with resistance across 15 isogenic strains. In vitro expression of two natural and two synthetic chimeric alleles shows that the Asp137 substitution alone is responsible for both the loss of E3's carboxylesterase activity and the acquisition of a novel OP hydrolase activity. Modeling of Asp137 in the homologous position in acetylcholinesterase suggests that Asp137 may act as a base to orientate a water mol. in the appropriate position for hydrolysis of the phosphorylated enzyme intermediate.

L6 ANSWER 39 OF 51 TOXCENTER COPYRIGHT 2003 ACS

DUPLICATE 23

ACCESSION NUMBER:

1997:57586 TOXCENTER

DOCUMENT NUMBER:

97364888 PubMed ID: 9221837

TITLE:

A physiologically based pharmacokinetic and

pharmacodynamic model for paraoxon in rainbow trout

AUTHOR(S):
CORPORATE SOURCE:

Abbas R; Hayton W L Division of Pharmaceutics and Pharmaceutical Chemistry,

College of Pharmacy, The Ohio State University, Columbus

43210-1291, USA

SOURCE:

TOXICOLOGY AND APPLIED PHARMACOLOGY, (1997 Jul)

145 (1) 192-201.

Journal Code: 0416575. ISSN: 0041-008X.

COUNTRY:

United States

DOCUMENT TYPE:

Journal; Article; (JOURNAL ARTICLE)

FILE SEGMENT: MEDLINE

OTHER SOURCE:

MEDLINE 97364888

LANGUAGE:

English
Entered STN: 20011116

ENTRY DATE:

Last Updated on STN: 20011116

Trout were exposed to an aqueous solution of 75 ng/ml paraoxon for 5 days at 12 degrees C. The relationships among paraoxon concentration in water and target organs, AChE inhibition, and carboxylesterase (CaE) detoxification of paraoxon were characterized quantitatively by development of a PBPK-PD model. The PKPD model structure consisted of brain, heart, liver, kidney, and remainder of the body, which were interconnected by blood circulation. The paraoxon tissue/blood partition coefficients were: plasma/water, 1.46; liver/plasma, 5.89; brain/plasma, 3.90; heart/plasma, 2.91; kidney/plasma, 0.45; and blood/plasma, 0.91. Turnover of AChE was characterized from a dose-response study, in which its zero-order synthesis rate and first-order degradation rate constant were determined in several tissues; for brain they were 7.67 pmol/min and 7.31 x 10(-5) hr(-1). The uptake and depuration clearances of paraoxon (Cl(u) = 0.651 and Cl(d) = 0.468 ml min(-1) g body wt(-1)) were determined using a compartmental model. During continuous water exposure to paraoxon, AChE activity in the tissues declined to new steady state values that were maintained by the synthesis of new AChE. CaE was shown by simulation to be an important pathway for detoxification of paraoxon.

L6 ANSWER 40 OF 51 EMBASE COPYRIGHT 2003 ELSEVIER SCI. B.V.

ACCESSION NUMBER: 97230850 EMBASE

DOCUMENT NUMBER:

1997230850 EMBA

TITLE:

Purification and characterization of guinea-pig liver

microsomal deacetylase involved in the deacetylation of the

O-glucoside of N-hydroxyacetanilide.

AUTHOR:

Suzuki-Kurasaki M.; Yoshioka T.; Uematsu T.

CORPORATE SOURCE: T.

T. Uematsu, Department of Chemical Hygiene, Hokkaido Inst.

Pharmaceutical Sci., Otaru 047-02, Japan

SOURCE:

Biochemical Journal, (1997) 325/1 (155-161).

Refs: 51

ISSN: 0264-6021 CODEN: BIJOAK

COUNTRY:

United Kingdom
Journal; Article

DOCUMENT TYPE: FILE SEGMENT:

029 Clinical Biochemistry

LANGUAGE: SUMMARY LANGUAGE: English English

A microsomal deacetylase that catalyses the deacetylation of the O-glucoside of N-hydroxyacetanilide (GHA) was purified from guinea-pig liver. The activity was located exclusively in the microsomes and not detected in the cytosol. The purified GHA deacetylase was a trimeric protein with a molecular mass of 160 .+-. 10 (S.D.) kDa composed of subunits of 53 .+-. 2 kDa; its pI was 4.7. The N-terminal amino acid sequence of GHA deacetylase was similar to those reported for guinea-pig and rat liver microsomal carboxylesterases. The GHA deacetylase showed a comparable hydrolytic activity towards p-nitrophenyl acetate (PNPA), although the activities towards N-hydroxyacetanilide, acetanilide and some endogenous acylated compounds were very low or not detectable. The deacetylase activity towards GHA was inhibited by organophosphates but not by p-chloromercuribenzoate, suggesting that GHA deacetylase can be classified as a B-esterase. The enzyme exhibited a positive homotropic cooperativity towards GHA. The values of the Hill coefficient, the half-saturating concentration ([S]0.5) for GHA, and V(max) were 1.59 .+~. 0.03, 5.51 .+~. 0.07 mM and 32.5 .+~. 1.4 .mu.mol/min per mg respectively, at the optimum pH of 8.5. The bell-shaped pH dependence of the V(max)/[S]0.5 profile indicated pK2 values attributed to histidine and lysine residues. The study of stoi-chiometric inhibition by di-isopropyl fluorophosphate and kinetic analysis with the Monod-Wyman-Changeux model suggests that GHA deacetylase has six substrate binding sites and three catalytically essential serine residues per enzyme molecule.